Black Carbon Content in Airway Macrophages is Associated with Reduced CD80 Expression and Increased Exacerbations in Former Smokers With COPD

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Citation: Tejwani V, Moughames E, Suresh K, et al. Black carbon content in airway macrophages is associated with reduced CD80 expression and increased exacerbations in former smokers with COPD. Chronic Obstr Pulm Dis. 2020; Published online November 6, 2020. doi: https://doi.org/10.15326/jcopdf.2020.0170

Running head: Macrophage black carbon content in COPD

List of abbreviations:

COPD – Chronic obstructive pulmonary disease

AM – Airway macrophages

CAT – COPD Assessment Test

mMRC – modified Medical Research Council Dyspnea Scale
ECSC – Ease of Cough and Sputum Clearance score

ATS – American Thoracic Society

Keywords: Macrophage, CD80, black carbon, COPD

Funding support:

Supported by grants from the National Heart, Lung, and Blood Institute (F32HL149258-01, V.T.), Baurenschmidt Award from the Johns Hopkins Eudowood Foundation (V.T.), Department of Defense (DoDW81XWH-16-1-0510, F.R.D.), National Institute on Minority Health and Health Disparities of the NIH (P50MD010431, N.N.H), the National Institute of Environmental Health Sciences of the NIH (R01ES022607, N.N.H), and the U.S. Environmental Protection Agency (EPA) (agreement number 83615001, N.N.H.). This manuscript has not been formally reviewed by the EPA or NIH, and the views expressed in this document are solely those of the authors and do not necessarily reflect those of the EPA or NIH.
Abstract:

Background: Chronic obstructive pulmonary disease (COPD) is characterized by recurrent exacerbations. Macrophages play a critical role in immune response and tissue repair in COPD. Airway macrophages (AM) are exposed to environmental exposures which are retained in the cytoplasmic material. Both biomass and particulate matter have been linked to higher AM black carbon. It is unknown if AM black carbon is associated with COPD morbidity and macrophage phenotype.

Methods: Former smokers with COPD were enrolled and sputum induction was performed to obtain airway macrophages. Macrophages underwent black carbon quantification and flow cytometry phenotyping. Health information was obtained the same day as sputum induction and prospective exacerbations were assessed by monthly telephone calls.

Results: We studied 30 former smokers with COPD that had a mean age of 67 years and mean FEV1% predicted of 60.8%. Higher AM black carbon content was associated with increased total exacerbations and severe exacerbations and reduced CD80 expression.

Conclusion: AM black carbon association with respiratory morbidity is largely unexplored and this is the first study to identify association with prospective exacerbations. Macrophages expressed reduced CD80, a surface marker providing costimulatory signals required for development of antigen-specific immune responses. Our findings suggest that reduced CD80 expression is the pathophysiologic mechanism for the association of AM black carbon content and increased exacerbations. Therefore, beyond solely serving as a marker for increased exposures, AM black carbon content may be predictor of future exacerbations given a macrophage less equipped to respond to an acute infectious exposure.
Background

Chronic obstructive pulmonary disease (COPD), the fourth leading cause of death worldwide,\(^1\) is characterized by recurrent exacerbations and progressive decline in lung function. Macrophages play a critical role in immune response and eventually tissue repair in COPD.\(^2\)–\(^4\) Macrophages from COPD patients compared to healthy controls exhibit reduced expression of surface markers important for immunity (adhesion molecules, antigen presentation molecules and recognition markers), as well as alterations in cytokine production.\(^5\)–\(^6\) Airway macrophages (AM) are also exposed to environmental exposures, such as cigarette smoke and pollution.\(^7\)

In developed countries, the most strongly supported cause of COPD is cigarette smoking; however, there is growing evidence that environmental exposures other than tobacco smoke, including indoor and outdoor air pollution, are linked to pulmonary morbidity and mortality.\(^8\),\(^9\) Inhaled fine particulate matter avoids upper airway mucociliary trapping and can reach the lower airways where airway macrophages (AM) reside. Subsequently, these particles are phagocytosed by AMs and retained in the cytoplasmic material. Both biomass and particulate matter exposure have been linked to higher AM black carbon.\(^10\),\(^11\) Whether black carbon content within AM influences COPD morbidity or macrophage phenotype is yet to be elucidated, therefore we sought to determine the relationship of black carbon content to COPD morbidity and macrophage phenotype.

Methods

Participant recruitment and macrophage carbon assessment

Former smokers with COPD were enrolled (\(n=30\)) after meeting the following inclusion criteria: age \(\geq 40\) years, post bronchodilator \(35\%<\text{FEV}_1\leq 80\%\) predicted, \(\text{FEV}_1/\text{FVC} < 70\%\), and \(> 10\) pack years smoking, with quit date of \(> 6\) months prior to enrollment and exhaled
carbon monoxide level $\leq 6$ ppm. Participants with evidence of active infection or exacerbation within two weeks of study visit did not undergo sputum induction and excluded from our study. Participants were followed longitudinally at baseline, 3 months and 6 months with comprehensive phenotyping and induced sputum collection. Informed written consent was obtained from all participants, all experiments were performed in accordance with relevant guidelines and regulations, and the study was approved under Hopkins IRB protocol number 00069904.

Sputum induction was performed according to the methods of Alexis et al. Briefly, the Aerosol Universal III nebulizer was used and sputum samples solubilized with Sputolysin and cytocentrifugation and cytospin slide generated with Diff-Quick for cell differentials. The pellet was re-suspended in freezing media and cryopreserved for future analysis. Digital color images of macrophages on cytospin slides were taken using the Nikon Eyepiece microscope camera (Nikon, Tokyo, Japan) at 100x magnification with oil immersion. Per sample, a total of 50 randomly selected macrophages with an intact cell wall were analyzed for black carbon content using ImageJ software and the mean carbon black area of macrophages was used for analysis. We also counted the fraction of the 50 macrophages that were positive for any black carbon and generated an independent variable of percent black carbon positive macrophages for a secondary analysis.

Health information was collected the same day as the sputum collection; specifically, the COPD Assessment Test (CAT), the modified Medical Research Council (mMRC) Dyspnea Scale and the Ease of Cough and Sputum Clearance (ECSC) score. Participants were also queried regarding their employment status and if currently working, asked if their current job exposed them to “vapors, gas, dust or fumes.” Patients were regarding the occurrence of mild,
moderate or severe exacerbations by monthly telephone calls. Mild, moderate and severe exacerbations were defined as worsening of symptoms leading to increase in inhaler use without antibiotics or systemic glucocorticoids, exacerbation leading to treatment with antibiotics or systemic glucocorticoids and exacerbation resulting in an urgent care visit, ED visit or hospitalization, respectively. Spirometry was performed according to American Thoracic Society (ATS) criteria.15

*Macrophage phenotyping by flow cytometry*

The first available sputum sample with at least 1 x 10^6 cells (n=16) was processed for flow cytometry. Cells were thawed and plated prior to incubation with Golgi Stop and Golgi Plug for 3 hours, then stained with Fixable UV Live/Dead, and then Fc blocked (BD). We stained with the surface markers BV515-conjugated anti-CD3 (UCHT1, BD Horizon), BB700-conjugated anti-CD19 (SJ25C1, BD Horizon), APC-Cy7-conjugated anti-HLA-DR (G46-6, BD Pharmingen), BUV395-conjugated anti-CD16 (3G8, BD Horizon) and BUV737-conjugated anti-CD14 (M5E2, BD Horizon), Pe-Cy7-conjugated anti-CD80 (L307.4,BD Pharmingen) and APC-R700-conjugated anti-CD274 (MIH1, BD Horizon). Cells were subsequently fix and permeabilized for 30 minutes and intracellular cytokines staining performed for 30 minutes followed by intracellular staining with APC-conjugated anti-IL-10 (JES3-19F1), BV421-conjugated anti-IL-1β (H1b-98, BioLegend), BV510-conjugated anti-IL-8 (G265-8, BD Horizon), PE-CF594-conjugated anti-TGF-β1(TW4-9E7), and BV605-conjugated anti-TNF-α (MAb11, BioLegend). Fluorescence minus one (FMO) was utilized to determine specificity for intracellular cytokine expression. Compensation was completed with UltraComp eBeads (eBioscience). Acquisition is performed using a FACSFlow instrument with FACSFlow software (BD) and FlowJo version 10.5.0 (Tree Star, Inc., Ashland, OR) for analysis. Flow cytometer
gating strategy will be as follows: single cells will be identified first, subsequently, a UV excitable live dead discrimination assay (Invitrogen) was applied. Live cells were excluded for CD3+ and CD19+ populations to remove T- and B-cells, respectively. After removing these other cell types, we characterized airway macrophages by gating those with characteristic side-scatter and forward-scatter properties and with CD14 positivity (Supplementary Figure 1).16 We then measured the mean fluorescence intensity (MFI) of each surface marker (HLA-DR, CD274 and CD80) and intracellular cytokines (IL-10, IL-1β, IL-8, TGF-β1, and TNF-α).

Analysis

Descriptive analyses were used to examine means and standard deviations or proportions and counts. We performed cross-sectional regression analysis using the data from the participant’s earliest visit for which he/she had induced sputum collection. Linear regression analyses were run to evaluate the relationship between AM black carbon content and COPD outcomes, and Poisson regression analyses for exacerbation risk. Models were adjusted for gender and inhaled corticosteroid use, defined as potential confounders based on their bivariate associations (p<0.2) with AM black carbon and any measured outcomes. Other potential confounders including total pack-years smoking and time since smoking cessation were considered but not included given they did not meet the criteria of p<0.2 bivariate association. Poisson regressions were performed taking into account follow-up days as offset and using robust standard error to account for potential overdispersion. For immune markers, given their smaller sample sizes and greater susceptibility to outlier influence, an additional analysis using Spearman’s rank correlation was conducted to test associations with AM black carbon. Statistical significance was defined as P<0.05, and, for immune marker analysis, Bonferroni-
corrected P-value was evaluated for multiple-comparisons. Analyses were performed with StataSE statistical software and R.

Results

Participant characteristics

Participants (Table 1) had a mean (±SD) age of 67 (±9.0) years, mean (±SD) pack-years of smoking of 51.7 (±36.9) with a mean (±SD) of 153 (±136.9) months since quit smoking, and a post-bronchodilator FEV1 % predicted of 60.8% (±12.8) predicted. The first available sputum was from baseline, and 3 and 6 month follow-up for 20, 5 and 5 participants, respectively. The samples that had adequate cell count for flow cytometry were obtained at baseline, and 3 and 6 month follow-up for 9, 3, and 4 participants, respectively. During the follow-up periods following sputum collection, 6 participants had one or more episodes of severe exacerbations, including one participant who reported three episodes during his/her time in the study. Fourteen participants had one or more episodes any exacerbation (mild, moderate, or severe) during their time in the study following sputum collection and the characteristics of these 14 are shown in Table 1. The average follow-up time following sputum collection was 189 days (~6 months) and the annualized rate of exacerbations was 2.1 for any exacerbation and 0.6 for severe exacerbations. Two participants did not have the data for prospective risk of exacerbations. Sixteen patients (53%) were using inhaled corticosteroids (Table 1) and no patients were on oral corticosteroids.

The mean (±SD) total cell count in the sputum was 45.6 x 10^5 with 24.5%±14 macrophages (Table 1). The mean sputum black carbon area was 3.78 µm². An example of images of airway macrophages exhibiting high and no black carbon loading is shown (Figure 1).

Association of airway black carbon with COPD outcomes
After adjustment for covariates, higher airway black carbon content was associated with increased rate of total exacerbations (Incidence Rate Ratio [IRR] = 1.20 per µm² increase, p=0.032) and severe exacerbations (IRR = 1.31, p=0.007), over the total follow-up period (Table 2). Similar results were identified in secondary analyses looking only at exacerbations in the three months following measurement. Airway black carbon content was not associated with other measured clinical outcomes: ECSC, FEV1, SGRQ, CAT or MRC, although directionally a higher black carbon content corresponded to worse outcomes for FEV1, SGRQ, and MRC (Table 2). Percent black carbon positive macrophages trended toward higher total exacerbations (IRR = 1.03 per percent increase, p=0.074) and was associated with prospective severe exacerbations (IRR = 1.05, p=0.02). Furthermore, percent black carbon positive was not associated with other clinical measures.

Flow cytometry

There were no significant differences between the participants that did and did not have adequate cell counts for flow cytometry (Table 1). Increased black carbon content in airway macrophage was associated with reduced CD80 expression in adjusted model (β = -3.41, P=0.007) but not in unadjusted model (β = -1.59, P=0.118), though Spearman rank correlation showed significant association in both unadjusted (ρSpearman=-0.57, P=0.019) and adjusted models (ρSpearman=-0.77, P=0.009) (Figure 2). However, with Bonferroni correction for multiple testing, these associations did not reach significance for adjusted models, P=0.062 (P_{Spearman}=0.072), respectively. There was no association with black carbon content and CD274, HLA-DR, IL-1β, IL-8, IL-10, TGF-β1, IL-1Ra and TNF-α expression. The results were similar when using Spearman’s rank correlations. Percent black carbon positive macrophages was
associated with reduced CD80 expression in adjusted model ($\beta = -0.51, P=0.018$), but not Spearman rank correlation ($\text{rho}_{\text{Spearman}}=-0.22, P=0.2$).

**Discussion**

Particulate matter and biomass exposure have been independently associated with macrophage carbonaceous loading\textsuperscript{10,11} and in separate studies, with worse COPD morbidity.\textsuperscript{17,18} However, it has not been established if the AM black carbon influences COPD morbidity or macrophage function. Here, we report the novel finding of AM black carbon association with prospective total and severe exacerbations among former smokers with COPD. Given the deposition of this carbonaceous material in the macrophage, we hypothesized that it will skew macrophage phenotype leading to an abrogated immune response as the cause of increased exacerbations. Through flow cytometry isolation of these cells, we identified the additional novel association of black carbon content with reduced AM CD80 expression proving a plausible mechanism for increased exacerbations.

Airway macrophage black carbon content is associated with environmental exposures,\textsuperscript{10,11,19,20} however its impact on respiratory disease morbidity has been largely unexplored. We hypothesized that the AM black carbon content would be associated with worse morbidity given dysregulated macrophage function. In the both the total follow-up period (an average of approximately six months) and the immediate next three months following assessment, increased AM black carbon content was associated with increased total and severe exacerbations. Although the exact lifespan of human AM is unknown, six-month exposures are most strongly associated with AM black carbon content\textsuperscript{20} and in a case report following bone marrow transplantation, patient’s AM disappeared linearly, with their life span being approximately 81 days.\textsuperscript{21} Therefore, the exacerbations captured in our follow-up period could
reflect the AM black carbon content we observed. In addition to the novel association of AM black carbon content with exacerbations, we identified directionally worse associations of AM black carbon content with other markers of COPD morbidity including a trend toward worse post-bronchodilator FEV1 % predicted ($\beta=-1.093, p=0.08$) suggesting we may have been underpowered to identify statistically significant associations with these outcomes. The trend toward a reduced FEV1 is consistent with what was identified by Kulkarni, et al. in a study of children with asthma.\textsuperscript{19} Notably, that study did not capture exacerbations and the association of AM black carbon content with exacerbations is previously unstudied. These findings have prompted interest in characterizing AM with increased black carbon content and speculation that their phagocytic ability may be impaired.\textsuperscript{22,23} We characterized these macrophages with the use of flow cytometry and identified AM black carbon content was associated with reduced CD80 expression.

The surface marker CD80 plays an important role providing costimulatory signals required for the development of antigen-specific immune responses.\textsuperscript{24} CD80 is also a marker for M1 macrophages which have enhanced phagocytic capability.\textsuperscript{25,26} Previous studies have demonstrated reduced macrophage CD80 expression in COPD subjects compared to healthy volunteers.\textsuperscript{27,28} Further supporting the importance of CD80 in COPD, increased serum autoantibodies against CD80 have been associated with worse COPD stage.\textsuperscript{29} Reduced CD80 can result in attenuated immunity, leading to a higher propensity for pathogen colonization which in turn is associated with advanced airway inflammation, increased frequency of exacerbation and accelerated decline in lung function.\textsuperscript{30–34}

There are limitations of this study that merit discussion. Our analysis is associative in nature and although our findings suggest a plausible biologic model, this alone does not prove
causation. Our sample size may be underpowered to detect the effect of AM black carbon content on other health outcomes, surface markers or intracellular cytokines. We utilized multiple surface markers and intracellular cytokines which poses the potential for spectral overlap however sought to mitigate this with the use of FMO and compensation beads; compensation performed using both FACSDiva software and FlowJo demonstrated no spectral overlap for any of the fluorophores. Although sample availability precluded this, future studies could purify CD80+ macrophages and measure black carbon content to further eliminate the possibility of spectral overlap influencing our findings. Additionally, more thorough macrophage phenotyping with additional surface markers (e.g. CD163, CD206) could be performed in future studies. We did not have a control group would also help identify if this relationship is unique to COPD. Lastly, an FEV$_1$ <35% predicted precludes safe performance of sputum induction and as such, this subset of more severe COPD patients was not included in our study. No participants in an exacerbation were enrolled which strengthens our findings given observations were made during steady state. Similar to others studies of sputum cell count COPD in steady state reporting over 70% neutrophils,\textsuperscript{35,36} we observed a neutrophil predominance although less than these studies.

Our findings suggest that reduction in CD80 expression is the pathophysiologic mechanism for the association of AM black carbon content and increased exacerbations in former smokers with COPD. Therefore, beyond simply serving as a marker for increased exposures, AM black carbon content may be predictor of future exacerbations given a macrophage less equipped to respond to an acute infectious exposure. This study furthers our understanding of the influence of AM black carbon content on COPD morbidity and macrophage phenotype.
Table 1. Participant characteristics

<table>
<thead>
<tr>
<th>Characteristics of</th>
<th>Characteristics of</th>
<th>Characteristics of</th>
</tr>
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<tbody>
<tr>
<td>All participant characteristics (n=30)</td>
<td>subset with exacerbation (n=14)</td>
<td>subset with adequate cells for flow cytometry (n=16)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>Mean ± SD or % (n)</th>
<th>Mean ± SD or % (n)</th>
<th>Mean ± SD or % (n)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>67.0 ± 9.0</td>
<td>67.5 ± 8.6</td>
<td>67.6 ± 10.3</td>
</tr>
<tr>
<td>Female %</td>
<td>53% (16)</td>
<td>57% (8)</td>
<td>69% (11)</td>
</tr>
<tr>
<td>Black %</td>
<td>47% (14)</td>
<td>43% (6)</td>
<td>38% (6)</td>
</tr>
<tr>
<td>Education</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Some college or above</td>
<td>60% (18)</td>
<td>71% (10)</td>
<td>50% (8)</td>
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<tr>
<td>Occupational status</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Employed</td>
<td>20% (6)</td>
<td>21% (3)</td>
<td>17% (1)</td>
</tr>
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<td>Current occupational vapor, gas, dust or fume exposure</td>
<td>6.7% (2)</td>
<td>0% (0)</td>
<td>0% (0)</td>
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<td>Unable to work</td>
<td>20% (6)</td>
<td>14% (2)</td>
<td>19% (3)</td>
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<td>Retired</td>
<td>43% (13)</td>
<td>43% (6)</td>
<td>50% (8)</td>
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<tr>
<td>Unknown</td>
<td>17% (5)</td>
<td>21% (3)</td>
<td>25% (4)</td>
</tr>
<tr>
<td>BMI</td>
<td>32.1 ± 6.3</td>
<td>30.6 ± 5.1</td>
<td>31.7 ± 6.1</td>
</tr>
<tr>
<td>normal</td>
<td>13% (4)</td>
<td>14% (2)</td>
<td>19% (3)</td>
</tr>
<tr>
<td>overweight</td>
<td>27% (8)</td>
<td>36% (5)</td>
<td>25% (4)</td>
</tr>
<tr>
<td>obese</td>
<td>60% (18)</td>
<td>50% (7)</td>
<td>56% (9)</td>
</tr>
<tr>
<td>Total pack-years</td>
<td>51.7 ± 36.9</td>
<td>51.3 ± 34.8</td>
<td>53.4 ± 40.9</td>
</tr>
<tr>
<td>Time since quit smoking (months)</td>
<td>153 ± 136.9</td>
<td>151.3 ± 147.2</td>
<td>158.8 ± 126.2</td>
</tr>
<tr>
<td>Use of ICS, % (n)</td>
<td>53% (16)</td>
<td>43% (6)</td>
<td>50% (8)</td>
</tr>
<tr>
<td>FEV1 % Pred.</td>
<td>62.1 ± 11.0</td>
<td>61.7 ± 10.2</td>
<td>61.0 ± 12.1</td>
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<tr>
<td>Sputum Inflammatory Cells, mean (SD)</td>
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<td></td>
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<tr>
<td>Total cell count x 10^5 cells / mL</td>
<td>45.6 ± 61.2</td>
<td>42.6 ± 44.7</td>
<td>45.9 ± 53.6</td>
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<td>Neutrophils, %</td>
<td>45.3 ± 23.8</td>
<td>49.6 ± 22.5</td>
<td>45.5 ± 27.8</td>
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<td>Macrophages, %</td>
<td>24.5 ± 14</td>
<td>19.7 ± 14.4</td>
<td>23.3 ± 15.0</td>
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<tr>
<td>Eosinophils, %</td>
<td>1.7 ± 2.5</td>
<td>2.1 ± 3.0</td>
<td>0.6 ± 0.9</td>
</tr>
<tr>
<td>Lymphocytes, %</td>
<td>1.5 ± 1.2</td>
<td>1.4 ± 1.1</td>
<td>1.4 ± 1.2</td>
</tr>
<tr>
<td>AM Black Carbon, µm^2 mean (SD)</td>
<td>3.8 ± 3.8</td>
<td>4.7 ± 4.0</td>
<td>3.7 ± 4.0</td>
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</table>
Table 2. AM black carbon content associations with COPD outcomes (n=30)*

<table>
<thead>
<tr>
<th></th>
<th>IRR</th>
<th>95% CI</th>
<th>p</th>
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<tr>
<td>Severe exacerbations over total follow-up period</td>
<td>1.309</td>
<td>1.075</td>
<td>1.593</td>
</tr>
<tr>
<td>Any exacerbation over total follow-up period</td>
<td>1.197</td>
<td>1.016</td>
<td>1.410</td>
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<tr>
<td>Ease of Cough &amp; Sputum (ECSC)</td>
<td>0.006</td>
<td>-0.339</td>
<td>0.352</td>
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<tr>
<td>SGRQ Total</td>
<td>0.849</td>
<td>-1.164</td>
<td>2.862</td>
</tr>
<tr>
<td>COPD Assessment Test (CAT)</td>
<td>0.092</td>
<td>-0.684</td>
<td>0.868</td>
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<tr>
<td>mMRC</td>
<td>0.091</td>
<td>-0.030</td>
<td>0.212</td>
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<tr>
<td>Post-bronchodilator FEV1 % predicted</td>
<td>-1.093</td>
<td>-2.329</td>
<td>0.144</td>
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</table>

*Adjusted by gender and ICS use

Figure 1. Digital color images of airway macrophages (AM) in induced sputum. Image (a) is an example of an AM with high content of intracellular black carbon and Image (b) has no black carbon content.

Figure 2. Black carbon content and relation to CD80 expression (MFI)

Supplementary Figure 1. Gating strategy for sputum macrophages
Acknowledgments:

Author Contributions: V.T., K.S., F.D. and N.N.H conceived of the presented idea. L.G. and K.R. recruited patients and performed sputum induction. V.T., E.M. and S.T. performed black carbon quantification. V.T., N.A. and F.D. performed flow cytometric and processed flow cytometry data. V.T., K.S., N.P., J.W., and N.N.H. performed biostatistical analysis. All authors discussed the results and contributed to the final manuscript.

Conflict of Interest Disclosures:

Drs. Tejwani, Suresh, Putcha, Alexis, Moughames, Tang, Romero, Woo, D’Alessio and Ms. Grammer have nothing to disclose. Dr. Hansel reports grants and personal fees from AstraZeneca, grants and personal fees from GSK, grants from Boehringer Ingelheim, grants from COPD Foundation, personal fees from Mylan, all outside the submitted work.
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Unprimed, M1 and M2 macrophages differentially interact with Porphyromonas

plasticity features of classical and alternatively activated human macrophages. *Am J


Figure 1
Figure 2
Supplementary Figure 1